# Mélanie GOULEY

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## PhD Candidate in genetic engineering

### **Education**

- 2019 2022 Strasbourg, France
  Trinational Biotechnology Engineering School (ESBS) - equivalent to M.Sc. in Engineering EUCOR Universities of Strasbourg, Freiburg I. Br. and Basel (France, Germany, Switzerland) Enzymology, Thermodynamics, Structural biology, Chemistry, Plant Physiology, Neurobiology, Genetic engineering, Immunology, Bioethics, Finances, Quality, Statistics, Molecular modelling, Process engineering, Plant bio-engineering, Programming.
- 2016 2019 Bachelor's degree in Life Sciences, speciality in physiology of organisms
- Caen, France Genetics, Biochemistry, Cellular biology, Immunology (in plants and animals), Microbiology, Cellular signalisation, Biostatistics, Populations et ecosystems.

### Work experiences

02.2023 -	ETH Zurich – PhD Candidate
present	"Precision gene editing and multi-omics profiling of patient-specific variants in human-derived stem cells
Basel, Switzerland	and neurons"
Switzenana	Supervised by Prof. Dr. Randall Platt and Prof. Dr. Barbara Treutlein
02-07.2022	Wellcome Trust Sanger Institute – Master's thesis
Cambridge,	Randomizing gene regulatory regions with prime editing
UK	Identified interesting enhancer regions by analysing genome-wide <b>chromatin data sets</b> and combining the findings with a literature search.
	Used <b>CRISPR prime editing</b> to insert multiple <b>recombinase recognition sequences</b> into enhancer clusters of these genes.
	Created cell lines with stable prime editor expression that achieve > 80% insertion efficiencies for a loxP site (these lines are now widely used by other people in the lab).
	Established <b>targeted Oxford nanopore sequencing with Cas9 enrichment</b> in the lab. Collaborated with a team at Imperial college to learn the method.
	Presented my work at 2022 International Mammalian Synthetic Biology Conference as a poster.
11-12.2021	Institute of Plants Molecular Biology CNRS – Master's project
Strasbourg,	Insertion of CRISPR/Cas9 prime editing complex into A. thaliana targeting AtmtPNPase gene
France	Validated an experimental process for the insertion of the prime editing complex into plants. Used Golden Gate and Gateway cloning methods for the generation of a binary vector and analysis of Sanger
	sequencing results.
	Performed <b>bacterial transformations</b> in <i>E.coli</i> and <i>A. tumefaciens</i> strains and <b>plant transformation.</b>
07-08.2021	TWISTAROMA & PCBIS – Intern in analytic chemistry
Strasbourg,	Analysis of secondary metabolic compounds in serum and in beer by GC-MS and LC-MS
France	Led a <b>project in collaboration</b> between the TWISTAROMA start-up and the PCBIS academic platform.
	Generated a protocol for the <b>derivation of non-volatile compounds by silylation</b> (TMS) for GC-MS analysis.
	Automated GC-MS samples preparation and analysis with <b>Gerstel MPS Autosampler</b> .
	<b>Created a library</b> recording non-volatile compounds detected in GC-MS after silylation. Compared compounds detection between <b>GC-MS and LC-MS.</b>
	compared compounds detection between <b>de-wis and re-wis</b> .

<b>06.2021</b> Strasbourg, France	EASE - Industrial bioproduction – Master's project Optimization of nanobodies production in Pichia pastoris (yeast) Production and optimization in baffled flasks. Production in 3L bioreactor in a clean room (GMP Class C). Filtration using a cross-flow filtration (TFF).
<b>2018 - 2019</b> Caen, France	University of Caen Normandy – <i>Teaching and Mentoring</i> Mentoring 1st year biology's students in different lectures. Weekly supervisions in groups of 4 to 5 students. Discussing and maintain a regular monitoring, preparing mock-exams.

## **Poster presentation**

M. Gouley, J. Koeppel, L. Parts (2022). "Randomizing gene regulatory regions using CRISPR/Cas9 Prime editing and the Cre Lox system". *Poster Session*. Edinburgh, UK – 2022 International Mammalian Synthetic Biology Conference.

#### **Skills**

Experimental	Genetic engineering: CRISPR/Cas9 Prime editing, Creation of stable cell line, Recombinases, Golden Gate cloning, Gateway cloning, Long-read sequencing. Molecular biology: PCR, Agarose Gel, Western Blot, DNA extraction, High Sensitivity DNA kit.
	<b>Cell culture:</b> K562, HAP1, HeLa, iPSCs, Neural differentiation, Transfections, Infection, Electroporation, Immunostaining, FACS, Biosafety Level 2.
	Chromatography: GC-MS, LC-MS, Ion exchanger (AKTA), Gel filtration.
Softwares	Benchling, ImageJ, FlowJo, IGV, Microsoft Projects, Familiarity with R.
Languages	French (Native), English (Fluent), German (Basic).
References	References available on request.